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Effects of Melanocortin 1 Receptor Agonists in Experimental Nephropathies

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Abstract

Nephrotic syndrome, characterized by massive proteinuria, is caused by a large group of diseases including membranous nephropathy (MN) and focal segmental glomerulosclerosis (FSGS). Although the underlying mechanisms are beginning to unravel, therapy is unspecific and far from efficient. It has been suggested that adrenocorticotropic hormone (ACTH) has beneficial effects in patients with MN and possibly in other nephrotic diseases. We have previously reported that ACTH may act directly on podocytes through the melanocortin 1 receptor (MC1R). In the present study, we evaluate the effect of highly specific MC1R agonists in two different nephrotic disease models. Experimental MN: Passive Heymann nephritis (PHN) was induced in rats that were treated for four weeks with MS05, a selective MC1R agonist, or saline. The degree of albuminuria was significantly reduced over time and the effect was sustained one week after treatment withdrawal (p < 0.05). Experimental FSGS: Based on a dose-response study, two doses of adriamycin were used for induction of nephropathy in Balb/c mice. Mice were treated with either a synthetic MC1R agonist (BMS-470539), with α-melanocyte stimulating hormone (α-MSH) or with saline. There was no beneficial effect of treatment. In summary, MC1R agonists reduce albuminuria and improve morphology in experimentally induced MN whereas they have no effect in experimental FSGS. The results illustrate the differences in these podocytopathies in terms of signaling mechanisms underlying proteinuria, and progression of disease.

Introduction

The clinical entity nephrotic syndrome is caused by different morphological diseases such as membranous nephropathy (MN), minimal change disease (MCD), membranoproliferative glomerulonephritis (MPGN), and focal segmental glomerulosclerosis (FSGS). Recent studies have partially revealed the molecular mechanisms underlying these diseases. Thus, antibodies against phospholipase A2 seem to be involved in the pathogenesis of MN [1] as those to megalin are in passive Heymann nephritis in rats [2]. For patients with primary FSGS, soluble urokinase receptor (suPAR) has been reported to be elevated [3]. However, many questions remain [4–7], and treatment options are still far from specific or effective [8].

It was previously reported that treatment with adrenocorticotropic hormone (ACTH) had beneficial effects in patients with a wide range of nephrotic diseases [9,10]. The effect was most prominent in patients with MN, but the treatment was also effective in some cases of MCD, diabetic nephropathy, MPGN and FSGS. The positive effects of ACTH on MN have been confirmed in a randomized controlled trial [11], albeit in low number of patients. In a recent study, we put forward a possible molecular mechanism behind the beneficial effects of the treatment [12]. Firstly, one of the ACTH receptors, melanocortin 1 receptor (MC1R), was expressed in the kidney, and more specifically in the podocytes. Secondly, MC1R agonists ameliorated the nephrotic disease in rats with passive Heymann nephritis (PHN), an experimental model of MN. Thus, ACTH and agonists of MC1R decreased proteinuria, reduced oxidative stress, and improved podocyte morphology.

Podocytes play a crucial role in the normal glomerulus and in the development of many glomerular diseases [13]. Although the onset of disease may vary, the resulting podocyte injury with foot process effacement and proteinuria is common in most proteinuric disorders such as human MN and FSGS. PHN in rats and adriamycin-induced nephropathy in mice are well-characterized models resembling human MN and FSGS respectively. PHN is characterized by immune deposits formed in situ in the glomerulus, triggered by antigen expression on the podocyte surface [14], mainly megalin [2]. Proteinuria reaches a peak level 14 days after disease induction and although immune deposit formation...
eventually ceases, proteinuria persists lifelong [14]. The adriamy-
cin model is characterized by glomerulosclerosis, tubulo-interstitial
inflammation and fibrosis [15,16]. Podocytes [17–19], as well as
endothelial cells and their surface layer [20], are damaged, leading
to proteinuria.

In this study, we used different MC1R agonists to treat two experi-
mental nephrotic diseases, MN and FSGS. The agonists used were:
MS05, a selective peptide agonist for MC1R [21];
BMS-470539, a synthetic highly selective MC1R agonist [22–24];
and α-melanocyte stimulating hormone (α-MSH), an agonist for
all melanocortin receptors 1–5 except type 2, which is selective
only for ACTH.

The aim of this study was to evaluate the effects of MC1R
agonists in two experimental nephrotic models, representing two
different morphological diseases. Our hypothesis was that MC1R
agonists would have similar beneficial effects in both MN and
FSGS, and that they would ameliorate proteinuria.

Materials and Methods

Experimental Protocol

Animals. All animals had free access to standard food and
water, and were housed in a room with a 12-hour dark-light cycle.
Anesthesia was induced and maintained by inhalation of isoflurane
(2–3% v/v, Schering-Plough, Stockholm, Sweden) mixed with air
(8 L/min) in an isoflurane vaporizer (Ohmeda Isotec 5, Simtec
engineering, Askim, Sweden). Temgesic® (0.1 ml/100 g body
weight, Schering-Plough, Stockholm, Sweden) was given as a post-
operative pain reliever in all small operative procedures. Gothen-
borg Ethical Board for Animal Experiments approved the study.

Experimental MN – Passive Heymann Nephritis

The experiments were performed on male Sprague Dawley rats
(Charles River, Germany) of initial body weight of 140–165 g. To
induce passive Heymann nephritis (PHN), Anti-Fx1A IgG
antibody, 30 mg/mL (Probetex Inc., San Antonio, TX), was slowly injected into the tail vein, 1.5 ml at day 0 and 0.5 ml at day
7. Instead of the PHN-inducing antibody, controls received sterile
saline. At day 14–15, treatment with MS05 (custom-made peptide
from Agridera, Vännäs, Sweden), 100 μg/day was started, either
via osmotic pump (Alzet® Osmotic Pumps, Cupertino, CA) placed
subcutaneously in the neck or via subcutaneous injections. The
osmotic pump was replaced after two weeks. Total treatment time
was four weeks. After two additional weeks, the rats were sacrificed
and the kidneys were harvested for further analyses. Weight was
followed twice a week. Spot urine samples were collected twice a
week and the analyses were based on 1–2 weekly observations per
animal: controls, n = 5 (7–10 observations/week); untreated PHN,
n = 14 (19–27 observations/week); MS05, n = 17 (19–33 observa-
tions/week).

Experimental FSGS – Adriamycin-induced Nephrotic Syndrome

All experiments were performed on male BALB/c mice
(Charles River, Germany) with an initial body weight of 22–
26 g. To establish a stable nephropathy, a dose-response study
using 5, 7, 8, 9 and 10 mg/kg (n = 5–19) was performed in order
to find the optimal doses of adriamycin. Treatment, either BMS-
470539 (30 μmol/kg, n = 18–19; synthetized by Enamine, Uk-
raine), or α-MSH (0.020 μmol/kg, n = 9; Sigma, St Louis, MO),
diluted in DMSO or 1:1 in water and PEG400 (Sigma, St Louis,
MO), was started one day before adriamycin was given, either via
osmotic pump (Alzet® Osmotic Pumps, Cupertino, CA) placed
subcutaneously in the neck or via daily subcutaneous injections.
Control mice, and mice given adriamycin with no subsequent
treatment, received vehicle only.

At day 0, FSGS was induced by a single tail vein injection of 8
or 10 mg/kg adriamycin (doxorubicin hydrochloride, SigmaAld-
drich, St Louis, MO). An equal volume of saline was given to con-
trols. To prevent weight loss due to low appetite, mice were
given one intraperitoneal injection of a 2 mL glucose-electrolyte
solution (16.7 g/L glucose in hypotonic 75 mM NaCl solution) on
day 1 to 11. Body weight was recorded and spot urine samples
were collected daily. At day 7 to 12, the mice were sacrificed and
the kidneys were harvested for further analyses.

Urinary Analyses

For rats, albumin was analyzed using the Rat Albumin Elisa
Quantitation Kit (Bethyl Laboratories Inc., Montgomery, TX) and
corrected for the urine creatinine concentration measured with the
Jaffé reaction using a creatinine standard solution (Sigma, St
Louis, MO). For mice, albumin was analyzed using the Albuwell
M ELISA Kit and all urine samples were corrected for the urine
creatinine concentration measured with the Creatinine Companion
Kit (both from Exocell Inc., Philadelphia, PA). Optical density
was measured on a Spectra max plus reader (Molecular Devices,
Sunnyvale, CA).

Blood Urea Nitrogen

Blood was taken from the mice through the caval vein at day 10
(8 mg/kg adriamycin) or day 8 (10 mg/kg adriamycin), centri-
fuged and plasma was immediately frozen and stored at −20°C.
Blood urea nitrogen (BUN) was measured with QuantiChrom™
Urea Assay Kit (BioAssay Systems, Hayward, CA) according to
manufacturer’s instructions.
Real-time PCR

RNA was prepared from sieved isolated glomeruli using the Qiagen Mini Kit with DNAse digestion (Qiagen Nordic, Solna, Sweden). RNA quality was confirmed using the Standard Sensitivity Kit on Experion™ (Bio-Rad, Hercules, CA). RNA was quantified using a NanoDrop spectrophotometer (NanoDrop Technologies Inc., Wilmington, DE). Reverse transcription of RNA was performed using the High Capacity RNA-to-cDNA kit at a total volume of 20 μL (Applied Biosystems, Foster City, CA). 50 ng of sample cDNA was used to quantify the mRNA level of each target gene by real-time PCR on the ABI Prism 7900 Sequence Detection system (TaqMan, Applied Biosystems (ABI), Foster City, CA), as previously described [25]. The following primers and probes, all verified by ABI, were used to detect mRNA of the following genes: GAPDH: Mm99999915_g1 (mouse), Rn01775763_g1 (rat), Hs99999905_m1 (human); MC1R: Mm00434851_s1 (mouse), custom made GenBankID AB306978.1 (rat), Hs00267167_s1 (human). All samples were run in duplicates. RNA samples without performing reverse transcription were used as negative controls. The comparative ΔΔCT method of relative quantification was used to calculate the differences in gene expression between the groups. GAPDH was used as endogenous control.

Electron Microscopy

Immediately after blood sampling and sacrifice, the kidneys were collected for morphological analysis. The renal artery and vein were clamped and the kidney was fixed by subcapsular injection of Karnovsky's fixative (2% paraformaldehyde and 2.5% glutaraldehyde in 0.05 M Na-cacodylate buffer, pH 7.2). The kidneys were cut into mm-slices and processed by standard procedure as previously described [26]. Slides were taken with a Leo 912AB Omega electron microscope (Leo Electron Microscopy Ltd., Cambridge, England) and examined in a blinded fashion by a pathologist as previously described [12]. Briefly, the number of podocyte foot processes per 10 μm glomerular basement membrane was determined in 2–5 places of each glomeruli, n = 15 glomeruli for all groups.

Western Blotting

Protein was prepared from isolated glomeruli and a previously described mouse podocyte cell line [27]. Protein from a human melanoma cell line, A375 (ATCC, Manassas, VA), served as a

Figure 2. MC1R agonists did not reduce albuminuria in adriamycin-treated mice. (A) Five different doses of adriamycin, in the range of 5–10 mg/kg, were intravenously injected and the level of albuminuria was measured after 7 days. The urinary albumin-to-creatinine ratio (UACR) increased with increasing levels of adriamycin. (B) Albuminuria at day 7. Treatment with the MC1R agonist BMS-470539 did not reduce the level of albuminuria for the 8 mg/kg adriamycin dose, on the contrary it slightly increased albuminuria for the 10 mg/kg dose (n = 17–19, p ≤ 0.05). The unspecific melanocortin receptor agonist α-MSH did not have any significant effect (n = 9, n.s.) compared with untreated adriamycin mice. Results are presented as mean ± SEM.

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Figure 3. Blood urea nitrogen (BUN) does not differ in untreated adriamycin mice compared with MC1R agonist treated mice. BUN was measured in plasma samples taken from mice 8–10 days after injection of adriamycin. There was no difference between the groups (n = 9–10, n.s.). Results are presented as mean ± SEM.

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positive control for MC1R expression. The protein samples were equally loaded (10 µg each) and western blotting was performed as previously described [12]. The following antibodies and dilutions were used for detection of specific proteins: anti-MC1R antibody 1:500 (AMR-620, Alomone Labs, Ltd., Israel) and anti-actin goat polyclonal antibody 1:1000 (sc-1615, Santa Cruz Biotechnology, Inc., Dallas, TX). An MC1R control peptide antigen (Alomone Labs, Ltd., Israel) was separately added at a 1:1 weight ratio to the experimental MN and there was no significant difference in the weight between the groups at any point of the experiment.

Mice with adriamycin-induced FSGS. Because of the narrow dose interval in adriamycin-induced nephropathy, a dose-response study was performed to ensure the optimal dose in this set-up. As can be seen in Figure 2A, the lowest dose of 5 mg/kg had UACR levels slightly increased compared to controls: 0.44±0.34 and 0.02±0.02, respectively. UACR was increased by the 7 mg/kg dose to 1.17±0.34, but with a large variation within the group and not high enough to ensure an established disease. Two of the higher doses were therefore picked for further treatment studies: 8 mg/kg, and 10 mg/kg.

To further explore the role of MC1R in nephritic disease, the highly selective, synthetic, and stable agonist BMS-470539, and the unspecific MC1R agonist a-MSH were used for treating adriamycin-induced nephropathy (Figure 2B). For 8 mg/kg there was no difference in the level of albuminuria compared with the untreated mice. Thus, untreated mice subjected to adriamycin had a UACR of 2.75±1.1 (n = 19), compared to mice treated with the MC1R agonist BMS-470539; 2.46±0.73 (n = 19, n.s.). For the higher adriamycin dose, 10 mg/kg, that induced a more severe disease, BMS-470539 treatment resulted in a slightly higher UACR, 13.2±2.8 (n = 18) compared with untreated adriamycin mice, 7.34±2.4 (n = 17, p<0.05). Treatment with a-MSH did not significantly increase UACR, 13.6±7.9 (n = 9, n.s.). Controls remained at normal levels, i.e. UACR <0.02 (n = 10). At day 12 after adriamycin injection, the level of UACR had increased further, but with no differences between the groups: untreated adriamycin mice, 12.2±1.9 (n = 7); BMS-470539, 16.7±2.4 (n = 9, n.s.); a-MSH, 15.9±3.2 (n = 6, n.s.).

Blood urea nitrogen (BUN) is commonly used to assess kidney function. As can be seen in Figure 3, there was no difference between mice with untreated adriamycin-induced nephropathy compared with those treated with BMS-470539 (n.s.).

Weight loss in adriamycin-treated mice was low due to daily intraperitoneal injections of a glucose-salt solution. At day 7, adriamycin-treated mice had lost approximately 5% of their initial body weight (p<0.05). There were no differences between the MC1R agonist treated groups and the vehicle-treated mice given adriamycin (n.s.).

Podocyte Morphology is Improved by MC1R Agonists in MN but not in FSGS

Treatment with the MC1R agonist MS05 improved morphology in the PHN rats [12]. Adriamycin treated mice displayed pathological changes with a focally disrupted glomerular barrier, including pronounced foot process effacement (Figure 4B). This effect was segmental and in line with the disease model, since FSGS nephrotic mice presented unaffected glomerular parts as well. Glomerular damage was also quantified by counting the number of foot processes along the glomerular basement membrane. Controls had 22.8±0.90 numbers of foot processes/
Glomerular Expression of MC1R in Rat and Mouse

We have previously shown with real-time PCR that MC1R mRNA is expressed in human and rat kidney tissue with a Ct level of around 28 [12]. In this study, glomerular MC1R gene expression in PHN rats was confirmed but, as can be seen in Table 1, the Ct levels were higher (33) reflecting lower expression in PHN rats was confirmed but, as can be seen in Table 1, the Ct levels were higher (33) reflecting lower expression specifically in the glomerular cell type called podocyte. The mRNA expression levels in mice glomeruli were analyzed with real-time PCR (Table 1). The gene was expressed at low levels in mice with a mean Ct level of 33. There was no change in expression between controls, MS05 treated and untreated PHN rats (n.s.).

Protein expression of MC1R was also confirmed in mouse by western blotting, both in glomerular tissue as well as in a mouse podocyte cell line (Figure 5). A375, a human malignant melanoma cell line A375. MC1R protein expression was detected at the expected size of 37 kDa (left panel).

Discussion

We have confirmed our previous findings that MC1R agonists can ameliorate passive Heymann nephritis mimicking human membranous nephropathy (MN). We also evaluated the role of MC1R in adriamycin-induced FSGS and found a different pattern: MC1R agonists did not reduce albuminuria in the FSGS-model, and did not improve podocyte morphology.

In our previous paper we showed that MC1R agonist treatment could ameliorate experimental membranous nephropathy in rats [12]. In the same study we demonstrated human MC1R mRNA expression in all glomerular cell types and MC1R protein expression specifically in the glomerular cell type called podocyte. Together, the human expression data and experimental data in rats propose a role for MC1R agonists in treating glomerular disease.

In this study, we have further evaluated the potential role of MC1R as a new treatment option.

In PHN rats treated with an MC1R agonist, the amelioration of albuminuria was sustained after treatment withdrawal, which is in line with clinical observations where albuminuria has been further reduced after removal of ACTH treatment [9,10,12]. These results suggest that the disease is reversible and that MC1R agonists affect the podocytes in a beneficial manner. Clinical data suggest that ACTH and melanocortin receptor agonists (MCRs) could ameliorate nephrotic disease [9–11]. Indeed, we have shown that MC1R co-localizes with synaptopodin, indicating specific expression in podocytes [12], which suggests a general role for the receptor in podocytopathic nephrotic diseases such as MN.

On the other hand, in experimental FSGS, specific MC1R agonists did not reduce albuminuria. There are possible explanations to this finding including different disease mechanisms between MN and FSGS, as well as species diversity in MC1R expression and function. The experimental models used mimic two different human glomerular diseases, with similar changes in podocyte appearance including loss of foot processes, but also with distinct morphological differences. In experimental FSGS, not only the podocytes, but also the glomerular endothelium, is dysfunctional [20] [29,29]. Thus, the glomerular endothelial cell surface layer is markedly reduced in thickness after the administration of adriamycin [20]. Indeed, the endothelial changes precede those in the podocyte [29]. In FSGS, the complex signaling between podocytes and endothelial cells, required for the maintenance of an intact glomerular barrier, seems to be disturbed [28]. In contrast, the pathophysiological mechanisms behind MN involve immune-complexes, antibodies to megalin in Heymann nephritis and PLA2R in humans. These differences may explain why MC1R agonists are more or less effective. Another explanation for the discrepancy between the two disease models could be that the adriamycin-induced FSGS-like disease develops rapidly, while clinical FSGS has a slower time course. We showed that MC1R is present in not just human kidney tissue but also in rat and mouse. The MC1R agonists are highly effective for the human MC1R. However, it has been reported that mouse MC1R does not respond as well to stimulation with some of these agents as human MC1R [30], and the homology between human and murine MC1R is only 76% [31]. Thus, several aspects need to be considered when interpreting diverse effects of MC1R agonists on albuminuria in the different experimental disease models.

In summary, MC1R agonists reduce albuminuria and improve morphology in experimentally induced MN whereas they have no effect in experimental FSGS. The results illustrate the differences in these podocytopathies in terms of signaling mechanisms underlying proteinuria, and progression of disease. Further investigations of the mechanism behind MC1R signaling may reveal new information of how these diseases develop and could be treated.

Figure 5. MC1R protein expression. Western blotting was performed in order to detect MC1R protein expression in mouse and human tissue. Left panel: incubation with a rabbit polyclonal MC1R antibody (Alomone Labs). Right panel: incubation with MC1R antibody and control blocking peptide antigen (BP) in order to show antibody specificity. A) mouse whole glomerular lysate, B) cultured wild type mouse podocytes and C) a human malignant melanoma cell line A375. MC1R protein expression was detected at the expected size of 37 kDa (left panel).

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References


Author Contributions

Conceived and designed the experiments: ALJ AG JE MEJ BH JN. Performed the experiments: ALJ AG JE BH JN. Analyzed the data: ALJ AG JE BH JN. Wrote the paper: ALJ AG MEJ BH JN.