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Number of Spikes in Climbing Fibers Determines the Direction of Cerebellar Learning

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Cerebellar learning requires context information from mossy fibers and a teaching signal through the climbing fibers from the inferior olive. Although the inferior olive fires in bursts, virtually all studies have used a teaching signal consisting of a single pulse. Following a number of failed attempts to induce cerebellar learning in decerebrate ferrets with a nonburst signal, we tested the effect of varying the number of pulses in the climbing fiber teaching signal. The results show that training with a single pulse in a conditioning paradigm in vivo does not result in learning, but rather causes extinction of a previously learned response.

Introduction

The prevailing view of cerebellar learning is that climbing fiber (CF) impulses induce synaptic modification in the cerebellar cortex, such as long-term depression (LTD) of parallel fiber to Purkinje cell synapses. A much studied model of cerebellar learning is eyeblink conditioning, where repeated presentations of a neutral conditional stimulus (CS), often a tone, followed by a reflex-eliciting unconditional stimulus (US), such as an air puff to the cornea, causes acquisition of a conditioned blink response (CR) (Gormezano et al., 1983). Eyeblink conditioning relies on the cerebellum (McCormick and Thompson, 1984), and acquisition of adaptively timed CRs requires the cerebellar cortex (Yeo et al., 1984; Hesslow and Yeo, 2002). During conditioning, Purkinje cells develop adaptively timed pause responses, “Purkinje cell CRs,” thought to be the physiological basis of learned eyeblinks (Jirenhed et al., 2007; Jirenhed and Hesslow, 2011a). Results from microstimulation experiments suggest that Purkinje cells receive information about the CS via the mossy/parallel fiber pathway (Steinmetz et al., 1985; Hesslow et al., 1999), and information about the US via the CFs (Mauk et al., 1986; Jirenhed et al., 2007).

Purkinje cell CRs can disinhibit the cerebellar nuclei and, in extension, generate motor output (Hesslow and Yeo, 2002). In addition, disinhibition of the cerebellar nuclei causes increased inhibition of the inferior olive (IO), via the nucleo-olivary pathway (Bengtsson and Hesslow, 2006). Thus, the Purkinje cell CR can potentially inhibit the IO and the US signal (Rasmussen et al., 2008).

Cells in the IO fire in high-frequency bursts (>250 Hz), typically consisting of one to six spikes (Armstrong and Rawson, 1979; Maruta et al., 2007; Mathy et al., 2009). Each spike elicits a corresponding EPSP in the Purkinje cell dendrite, but only a single complex spike will appear in extracellular recordings. Thus, additional spikes do not result in extra complex spikes, although they alter the appearance and duration of the complex spike (Mathy et al., 2009).

The burst nature of the IO response was discovered decades ago (Eccles et al., 1966), but virtually all research on cerebellar learning, in which climbing fibers were stimulated, has used single pulses as teaching signals, assuming they are of an “all-or-none” nature (Ito, 2001). Recently, the nature of the CF signal has received renewed interest and the bursting nature of the IO response has been reaffirmed several times (Maruta et al., 2007; Mathy et al., 2009).

Here we tested the effect of varying the number of climbing fiber stimulus pulses on the acquisition and extinction of Purkinje cell CRs. A burst-like stimulus reliably results in acquisition of Purkinje cell CRs. Here we show that single climbing fiber impulses do not result in learning, but actually cause extinction of previously acquired Purkinje cell CRs.

Materials and Methods

Surgery: Thirteen male ferrets were anesthetized in a mixture of O2 and air with 1.5–2% isoflurane (Baxter Medical), which was subsequently replaced intravenously by propofol. During anesthesia, a tracheotomy was performed, and the gas was led directly into a tracheal tube. The end expiratory CO2 concentration, arterial blood pressure, and rectal temperature were monitored and kept within physiological limits throughout the experiment. During the whole experiment, infusion was given intravenously [50 mg/ml glucose and isotonic acetate Kinger’s solution (proportion, 1:1) with 0.004 mg/ml albumin fraction V (from bovine serum; Sigma-Aldrich), 2 mg/kg/h]. After fixation of the head in a stereotaxic frame, the skull was opened on the left side, and the caudal half of the left cerebral hemisphere, together with a substantial part of the...
Stimulation. Climbing fibers were stimulated in the ipsilateral inferior cerebellar peduncle by lowering an electrode 4.0–5.0 mm below the posterior cerebellar surface, at an angle of 45°, 4 mm lateral to the midline and 4 mm rostral to the caudal border of the vermis. Mossy fibers in the ipsilateral middle cerebellar peduncle were stimulated with an electrode lowered into the brainstem through the inferior colliculus at an angle of 80°, 1 mm medial to the lateral border of the inferior colliculus and at a depth of 4.0–5.0 mm. The superior colliculus was stimulated by an electrode lowered 100 μm below the surface of the middle of the superior colliculus at a 90° angle. While tracking for climbing fibers and mossy fibers, single stimulus pulses were applied, and field potentials were recorded in the C3 zone of hemispheral lobule VI (HVI), identified by previously established criteria (Hesslow and Ivarsson, 1994; Hesslow, 1994; Fig. 1B). Direct stimulation of cerebellar afferents was done with tungsten electrodes (diameter, 30 μm; deinsulated tip, 50 μm). The effectiveness of all stimulation sites and stimulus intensities were verified again and adjusted if necessary, when recording the activity of single Purkinje cells.

Training protocol. The CS consisted of a 300 ms, 50 Hz train of stimuli applied to the ipsilateral forelimb skin (1.5–2.0 mA, 0.5 ms), mossy fibers, or the superior colliculus (in both cases, 90–200 μA, 0.1 ms). Before the switch to a single pulse was made, the US consisted of two 10 ms stimulus trains delivered with a 20 ms interval, each consisting of five impulses at 500 Hz, applied to the ipsilateral climbing fibers (15–90 impulse trains delivered with a 20 ms interval, each consisting of five impulses at 500 Hz). A burst-like stimulus was used because it better elicits EPSPs in Purkinje cell dendrites than single-pulse CF stimuli. If Purkinje cell CRs were absent, we resumed training with the CS paired with a single-pulse CF stimulus. If Purkinje cell CRs were present, we resumed training with the CS paired with a burst-like US. In intracellular recordings, which rarely lasted >60 s, the protocol consisted of one, two, or three stimulus pulses applied to the climbing fibers at 500 Hz, with 1 s between each burst.

Results

The decerebrate ferret preparation (Fig. 1A) has several advantages when studying the mechanisms of eyeblink conditioning. Though the cell recordings presented here are from paralyzed animals, decerebrate ferrets can acquire conditioned eyelinks (Hesslow et al., 1999; Hesslow and Yeo, 2002), meaning that the...
neural network responsible for this learning remains intact. The relative stability of the setup allows for long single-unit recordings, which are necessary to follow the acquisition or extinction of Purkinje cell CRs. In addition, anesthesia can be excluded.

When we first tested IO/CF stimulation as a teaching signal, we used a protocol analogous to that which induces learning with a periorbital US (three pulses; 100–300 μA; 20 ms pulse interval, 50 Hz; 0.1 ms pulse duration). Although we always verified that CF stimulation elicited complex spikes in the Purkinje cell, this training protocol, with a nonburst CF stimulus, failed to induce any Purkinje cell CRs in nine consecutive experiments. To ensure that single CF pulses were indeed transmitted to the Purkinje cells and that these results not merely reflected insufficient signal strength, we used micropipettes to record EPSPs in Purkinje cell dendrites following one, two, or three CF stimulus pulses. The results confirmed that the number of EPSPs elicited corresponded to the number of CF stimulus pulses applied (Fig. 1C), which is also consistent with previous research (Mathy et al., 2009).

After these nine failed attempts to obtain acquisition of Purkinje cell CRs, we switched to a teaching signal that more resembled normal IO firing (Eccles et al., 1966; Maruta et al., 2007; Mathy et al., 2009), namely, two 500 Hz bursts with five stimuli in each. This reliably resulted in acquisition of Purkinje cell CRs (Jirenhd et al., 2007). Subsequently, nearly all Purkinje cells (more than 100 cells), provided they had input from the relevant climbing fibers and mossy/parallel fibers, acquired Purkinje cell CRs when using climbing fiber burst stimulation as the US.

Motivated by the observation that two sets of five CF pulses at 500 Hz resulted in learning, whereas three pulses at 50 Hz did not, we designed an experiment to test how the number of CF pulses in the US affects the acquisition and extinction of Purkinje cell CRs. Animals were conditioned with a US consisting of two sets of five CF stimuli at 500 Hz (200–700 μA), delivered to the CF, paired with a CS consisting of a 300 ms pulse train at 50 Hz, delivered to the ipsilateral forelimb (n = 8; 0.6–1.5 mA; 1 ms), mossy fibers (n = 3; 100 μA; 0.1 ms), or superior colliculus (n = 1; 150 μA; 0.1 ms). When a Purkinje cell CR had been acquired, we switched from a burst to a single-pulse climbing fiber stimulus. Other variables were kept constant.

Fifteen sessions from 13 cells were recorded using a single-pulse CF stimulus after Purkinje cell CRs had been acquired. On three occasions, the recording was sufficiently long lasting to allow us to later switch back to a burst US, and then to a single pulse again. In 12 of 15 sessions, training with a single pulse CF stimulus resulted in extinction of the Purkinje cell CR (an increase in simple spike firing during the CS; Fig. 2). In two sessions there was no change in simple spike activity, and in one session there was a decrease in simple spike activity. The median change in simple spike activity was 41%, ranging from −17 to 116% (Fig. 2F). A Wilcoxon matched-pairs signed rank test confirmed that the simple spike frequency increased significantly during training with a single pulse (W = 104; p = 0.0015). Consistent with previous findings (Jirenhd et al., 2007; Jirenhd and Hesslow, 2011a,b), and with behavior (Gormezano et al., 1983), the de-
increase and subsequent increase in simple spike firing was most pronounced toward the end of the CS (Fig. 2E).

The US threshold for induction of a Purkinje cell CR was defined in one Purkinje cell initially trained with a forelimb CS (50 Hz, 1 mA), paired with two sets of five 500 Hz CF pulses (230 μA), until a Purkinje cell CR had been acquired. As expected, a single-pulse US caused extinction of the Purkinje cell CR. Increasing the number of pulses to two (500 Hz) did not result in reacquisition. However, three pulses (500 Hz) did result in reacquisition, and switching back to one pulse caused reextinction of the response (Fig. 3). This suggests that the equilibrium level is close to two spikes.

Occasionally, climbing fiber stimulation elicited simple spikes, possibly because of antidromic activation of mossy fiber collaterals. However, the rate of acquisition appears to be independent of this increase in simple spikes. Burst activation of climbing fibers or the inferior olive consistently results in Purkinje cell CRs, even if no simple spikes are elicited (Jirenheid et al., 2007). It is therefore unlikely that this increase in simple spike firing affected the present results.

Discussion

We have demonstrated that a US, or teaching signal, consisting of a single climbing fiber stimulus causes extinction, not acquisition, of a Purkinje cell CR. This was not because the stimulus failed to activate the climbing fibers since we always confirmed that the CF stimulus elicited a complex spike in the Purkinje cell. These results challenge the interpretation of research on cerebellar learning where an all-or-none teaching signal has been assumed (Ito, 2001).

For example, several studies have suggested that LTD, which is induced by pairing parallel fiber stimulation with CF stimulation, is the synaptic mechanism responsible for classical conditioning (Ito, 2001; Koekkoek et al., 2003). However, a single CF impulse, which has been shown to be sufficient for LTD induction (Ito et al., 1982; Ekerot and Kano, 1985; Ito, 2001), actually causes extinction of Purkinje cell CRs. These results, together with additional physiological evidence (Hesslow et al., 2013) as well as evidence from transgenic mice (Schonewille et al., 2011), suggest that LTD is not the critical mechanism for conditioning, although it is still possible that LTP plays an important role (Schonewille et al., 2010). In contrast to the LTD studies cited above, Mathy et al. (2009) observed potentiation of the parallel fiber–Purkinje cell synapse when pairing parallel fiber stimulation with a single CF impulse. Only when a CF burst was used did they get LTD. These recent discoveries, and the results presented in the present paper, raise some intriguing questions concerning the plastic changes underlying cerebellar learning. To what extent are depression and potentiation of the parallel fiber–Purkinje cell synapses responsible for the acquisition of Purkinje cell CRs, and for cerebellar learning in general? If depression and potentiation, which are induced within minutes of applying the stimulation protocol, are the underlying mechanisms of cerebellar learning, then why does acquisition of conditioned eyeblinks and Purkinje cell CRs take several hours? How exactly does the number of spikes in the climbing fiber signal influence the type of plasticity induced in the cerebellar cortex? These important questions should be addressed in future studies.

The reason that single CF impulses are insufficient to induce cerebellar learning may have to do with the fact that the IO serves multiple purposes. One function of the IO, besides relaying the teaching signal, is to regulate background activity in the cerebellar circuit (Cerminara and Rawson, 2004; Bengtsson and Hesslow, 2013). Interfering with the IO’s normal firing pattern has striking effects on cortical activity and, consequently, on cerebellar output (Bengtsson et al., 2004; Zbarska et al., 2007). Thus, to maintain normal functioning of the cerebellar network, it may be advantageous if modulation of learning only requires slight alterations of IO activity.

Furthermore, if a periocular stimulus can generate a burst of CF spikes (Fig. 1C), then the Purkinje cell CR, through inhibition of the IO, could reduce the number of CF spikes and consequently the number of EPSPs generated by the US. Thus, the negative feedback exerted by the Purkinje cell CR on the IO (Hesslow and Yeo, 2002; Bengtsson and Hesslow, 2006; Rasmussen et
al., 2008) may be able to push the strength of the US signal below the learning threshold. This would allow the cerebellar circuit to reach an equilibrium where the negative feedback from the Purkinje cell CR matches the strength of the teaching signal, thus preventing runaway synaptic plasticity in the cortex (Lepora et al., 2010). It also would explain why lowering the US intensity can lead to extinction of overt CRs (Kehoe and White, 2002). As long as the negative feedback generated by the Purkinje cell CR matches the strength of the US signal, neither acquisition nor extinction ought to occur. However, if the US intensity is reduced, the Purkinje cell CR could suppress the US signal below the learning threshold, causing extinction.

Placing our results in a broader context invites comparisons with learning in the hippocampus, where it has been known for a long time that opposite forms of plasticity can be induced by slight alterations of stimulus parameters (Malenka and Bear, 2004). In a similar manner, and consistent with the nonlinear relationship between cerebellar plasticity and calcium release in Purkinje cells (Vogt and Canepari, 2010), our results demonstrate that the exact stimulus parameters used determines whether you get acquisition or extinction of the Purkinje cell CR.

**References**


